

# 1 Why does the microbiome affect behaviour?

2 Katerina V.-A. Johnson<sup>1</sup> and Kevin R. Foster<sup>2</sup>

3 1. University of Oxford, Department of Experimental Psychology, South Parks Road, Oxford OX1  
4 3UD, UK

5 2. University of Oxford, Department of Zoology and Oxford Centre for Integrative Systems  
6 Biology, Oxford OX1 3PS, UK

7 Correspondence to K.R.F. [kevin.foster@zoo.ox.ac.uk](mailto:kevin.foster@zoo.ox.ac.uk) or K.V.-A.J. [katerina.johnson@pmb.ox.ac.uk](mailto:katerina.johnson@pmb.ox.ac.uk)

8

9

## 10 Abstract

11 There is growing evidence that the mammalian microbiome can affect behaviour and several  
12 symbionts even produce neurotransmitters. One common explanation for these observations is  
13 that symbionts have evolved to manipulate host behaviour for their benefit. Here we evaluate  
14 the manipulation hypothesis by applying evolutionary theory to recent work on the gut-brain  
15 axis. Although the theory predicts manipulation by symbionts under certain conditions, these  
16 appear rarely satisfied by the genetically-diverse communities of the mammalian microbiome.  
17 Specifically, any symbiont investing its resources to manipulate host behaviour is expected to be  
18 outcompeted within the microbiome by strains that do not manipulate and redirect their  
19 resources into growth and survival. Moreover, current data provide no clear evidence for  
20 manipulation. Instead, we show how behavioural effects can readily arise as a by-product of  
21 natural selection on microorganisms to grow within the host, and natural selection on hosts to  
22 depend upon their symbionts. We argue that understanding why the microbiome influences  
23 behaviour requires a focus on microbial ecology and local effects within the host.

24

## 25 Introduction

26 The link between the gut and brain has been discussed for centuries, with multiple proposed  
27 mechanisms underlying this relationship<sup>1</sup>. These include communication through the vagus  
28 nerve<sup>2</sup>, the immune<sup>3</sup> and endocrine systems<sup>4</sup> and microorganism-derived neuroactive  
29 chemicals<sup>5</sup>. The relative importance of these routes, and how they might interact, is unclear but  
30 studies are increasingly documenting effects of gut microorganisms on the brain and behaviour<sup>6–</sup>  
31 <sup>9</sup>. To describe these relationships the term ‘microbiota-gut-brain axis’ has been coined<sup>10</sup>. For  
32 example, faecal microbiota transplantation in mice can cause behavioural traits of the recipient  
33 to become more like those of the donor<sup>11</sup>. Behavioural effects have also been traced to specific  
34 subsets of the microbiota. There is evidence that *Lactobacillus* and *Bifidobacterium* species can

35 alleviate anxiety and depressive-like symptoms<sup>12-17</sup>, including in humans<sup>18,19</sup>. Particular  
36 *Lactobacillus* species can also improve social interactions in stressed mice<sup>20</sup> and restore impaired  
37 oxytocin production and social deficits driven by a high-fat maternal diet<sup>21</sup>. In addition,  
38 *Bacteroides* species have been shown to ameliorate repetitive and anxiety-like behaviours and  
39 communicative impairments in mice, seemingly through restoration of a specific bacterial  
40 metabolite<sup>22</sup>.

41 Such microbiota-driven alterations of host behaviour have led to the hypothesis that  
42 some symbionts manipulate the host for their own ends<sup>23-26</sup>. For example, microorganisms might  
43 make us more sociable<sup>27</sup>, and even altruistic<sup>28</sup>, in order to increase host contacts and enhance  
44 their transmission. The general idea of behavioural manipulation – whereby a microorganism  
45 evolves to change host behaviour because this increases microbial fitness<sup>29</sup> (for example,  
46 promoting its own transmission) – has its roots in parasitology<sup>30</sup>. Numerous parasites affect the  
47 host nervous system and drive atypical behaviour (Box 1), often by interfering with  
48 neurotransmitter or neuropeptide signalling<sup>31,32</sup>. These effects are commonly attributed to  
49 parasite manipulation<sup>30</sup>. Many examples come from invertebrate hosts and one particularly  
50 striking example is the fungus *Ophiocordyceps unilateralis* that infects insects, including ants. In  
51 this example, two key pieces of evidence support evolved parasite manipulation. Firstly, infection  
52 with the fungus induces hosts to adopt a certain elevation in the canopy and then bite on  
53 vegetation, anchoring the ant prior to fungal sporulation<sup>33,34</sup>. Secondly, and critically, there is  
54 evidence that this provides fitness benefits to the parasite, as the particular elevation (and likely  
55 humidity) that the ant adopts appears important for fungal development<sup>33</sup>. In mammals,  
56 examples of parasites that affect the behaviour of their hosts are rare but include rabies virus  
57 and *Toxoplasma gondii*<sup>35-37</sup>. However, although there is evidence that parasites can influence  
58 host behaviour, the second step – that the parasite gains a fitness benefit from its effects – is  
59 difficult to demonstrate. This is because it is typically challenging to show that the change in host  
60 behaviour resulting from parasite infection enhances the parasite's fitness in its natural  
61 environment. Thus, even in the parasite field, it is unclear whether some long-discussed examples  
62 of parasite manipulation are genuine<sup>38</sup>.

63 The parasite literature, therefore, teaches us that demonstrating evolved manipulation is  
64 experimentally challenging. This literature also makes evolutionary predictions about when one  
65 should expect host manipulation. Here we apply the evolutionary theory of parasite  
66 manipulation<sup>39,40</sup> and host-symbiont interactions<sup>41</sup> (Box 2) to the mammalian microbiome (Fig.  
67 1). We consider the possible routes by which natural selection may have led to host manipulation  
68 by gut microorganisms, and conclude that manipulation of host behaviour is often unlikely (in  
69 contrast to local modification of the gut environment). We explore other evolutionary  
70 explanations for the behavioural effects of mammalian symbionts and propose that they  
71 modulate behaviour as a side effect of natural selection on other functions. In particular, host-  
72 affecting compounds can arise as a by-product of natural selection on microorganisms to  
73 compete within or control the local environment. Finally, hosts may evolve to depend upon

74 microbial metabolites for normal physiological function and so if the microorganism that  
75 produces these metabolites is lacking, behavioural dysfunction can result.

76

### 77 **Manipulation of host behaviour**

78 The potential benefits to a symbiont from manipulating host behaviour, which we define here as  
79 global manipulation (Fig. 1), are clear; how a host behaves can strongly affect the growth and  
80 survival of a symbiont and its transmission to other hosts. Despite this, the conditions that favour  
81 the evolution of a manipulative trait are quite restrictive<sup>39,40</sup> (Table 1). Consider a bacterial strain  
82 that uses a dedicated set of enzymes to generate a compound that affects host behaviour.  
83 Moreover, let us assume that this compound influences host behaviour in a way that benefits the  
84 bacterium. Hypothetically, it could immediately make the host more sociable and increase the  
85 potential routes of transmission to new hosts, or it might make a host consume resources that  
86 the bacterium needs. When will the production of this compound be favoured by natural  
87 selection? If the host is only colonised by this single strain, production of the compound is  
88 predicted to evolve so long as any fitness cost of production is outweighed by the benefits of  
89 increased nutrients or transmission.

90 If the bacterium must compete for resources and space with other strains and species,  
91 however, the prediction is very different. Whereas the metabolic cost of the enzymes falls on the  
92 producing bacterium, the benefits are now shared by multiple members of the microbiota.  
93 Indeed, in the case of a transmission effect, it is likely that much of the microbiota benefit. If a  
94 bacterial strain manipulates host food preference, only strains in a similar niche to the producing  
95 strain may benefit, but these are also the main competitors of this strain. In either of these cases  
96 of hypothetical manipulation, therefore, the prediction is that a strain in the same niche that  
97 lacks the enzymes will outcompete the producing strain, because it receives the benefit without  
98 paying the cost. This competitor could be a loss-of-function mutant of the producing strain or  
99 another species that inhabits the same niche. Ultimately, this is predicted to lead to the loss of  
100 the manipulating compound<sup>39,40</sup>. For high costs of production, this loss is expected to happen  
101 rapidly, in a few microbial generations. For low costs, the loss is predicted to take longer. Low  
102 costs are possible because natural selection is expected to minimise the costs associated with a  
103 given trait for a given level of benefit<sup>42</sup>. Consistent with this, some parasites of invertebrates are  
104 thought to act by increasing synthesis of neuromodulators such as serotonin by the host, which  
105 may be less costly than producing it themselves<sup>31,43</sup>. A low cost may also be facilitated when a  
106 microorganism can use pre-existing metabolic pathways to drive host effects. However, even for  
107 low metabolic costs, the prediction is that a manipulative trait will eventually be lost in the face  
108 of prolonged competition from a non-producing strain within a host.

109 Evolutionary theory then predicts that the evolution of manipulation will critically depend  
110 on the diversity within the microbiota and, more specifically, how much competition a given  
111 strain experiences with other genotypes in its niche. If a strain is largely free from such

112 competition, manipulation is predicted to evolve if affecting host behaviour can increase  
113 resources or transmission. However, when a strain faces competition from other genotypes, the  
114 evolution of costly mechanisms of manipulation is disfavoured, as these will undermine the  
115 ability of a strain to persist in the microbiota. The question then is which of these two scenarios  
116 best represents a given host microbiome. The human gut is an ecologically complex community,  
117 estimated to contain hundreds to thousands of interacting species and strains<sup>44,45</sup>. Moreover,  
118 there is growing evidence for the importance of direct competition between strains of the same  
119 species, and between species. This derives both from ecological modelling<sup>46</sup> and empirical  
120 estimates of species interactions in the mouse gut<sup>47,48</sup>, and also from studies revealing the key  
121 role of bacteriocins and type VI secretion systems for ecological success in the gut<sup>49-51</sup>. These  
122 competitive conditions are predicted to lead to natural selection against symbionts – whether  
123 they are mutualists, parasites or commensals – that manipulate host behaviour (Table 1, Box 3).

124 How do our predictions relate to current discussions of host manipulation by the  
125 microbiota? In contrast to our predictions, a recent theory paper proposed that social  
126 (specifically altruistic) behaviour in animals can be explained by the evolution of microbial  
127 manipulation<sup>28</sup>. However, unlike the models of parasite manipulation<sup>39,40</sup>, this study simply  
128 assumed there was no microbial competition within hosts to disfavour a manipulating strain. It  
129 does not, therefore, challenge our predictions. If the microbial competition that occurs within  
130 the gut were accounted for in this microorganism-induced altruism model<sup>28</sup>, the expectation is  
131 still natural selection against manipulation<sup>39,40</sup>. There is, nevertheless, empirical evidence  
132 consistent with host manipulation by *Salmonella enterica* serovar Typhimurium, mediated by the  
133 vagus nerve<sup>52</sup>. Bacterial infections can trigger sickness-induced behaviour in hosts<sup>53,54</sup>. It is  
134 challenging to demonstrate that sickness behaviour is an adaptation to combat infection rather  
135 than a by-product of compromised physiology. However, one feature – loss of appetite – may  
136 function to decrease nutrient supply to intestinal pathogens<sup>53</sup>. *S. Typhimurium* seems to suppress  
137 this appetite loss, which may represent manipulation of host feeding behaviour in order to  
138 counteract a potential reduction in nutrient supply<sup>52</sup>. Curiously, in this case, this effect appeared  
139 to improve host fitness as well as microbial fitness, though estimates of host fitness in a  
140 laboratory system may not capture fitness effects seen in the natural environment. In any case,  
141 the data appear consistent with the evolution of microbial manipulation of behaviour.

142 Does this *S. Typhimurium* example contradict our predictions? Consideration of *S.*  
143 *Typhimurium* biology suggests not; rather this example is consistent with the predictions of when  
144 true manipulation can evolve. The evolutionary success of *S. Typhimurium* is based on its ability  
145 to transiently outcompete other species and become dominant in the gut<sup>55,56</sup>. This competitive  
146 dominance means that *S. Typhimurium* is not expected to be outcompeted by other species that  
147 do not invest in manipulation (Table 1). There remains the potential for a non-manipulating strain  
148 of *S. Typhimurium* to outcompete a manipulating one. However, this outcome would require  
149 simultaneous co-infection with multiple strains to be common, yet multiple-strain infections are  
150 rare for bacterial pathogens<sup>57</sup>. Importantly, the ecology of *S. Typhimurium* contrasts with the  
151 typical ecology expected in the gut microbiota. Many species exist at relatively low frequency

152 and face competition from other strains and species over long periods, spanning many symbiont  
153 generations. These conditions are well captured by the theory<sup>39,40</sup>, with the expectation that  
154 manipulation will often be disfavoured. Many species in the microbiota are likely to experience  
155 long-term competition, including members of the genera *Bifidobacterium* and *Lactobacillus* that  
156 are most associated with effects on host behaviour<sup>6-9</sup>.

157

### 158 **Local manipulation in the host**

159 We have discussed how the manipulation of global host phenotypes, such as behaviour, is only  
160 expected under specific conditions. However, there is growing evidence that the microbiota can  
161 affect host behaviour. What then are the alternative explanations for these effects of microbial  
162 symbionts on host behaviour? One explanation is that symbionts are naturally selected to  
163 manipulate the local gut environment, and this then influences host behaviour as a side effect.  
164 Local manipulation is predicted to be more likely than behavioural manipulation because there  
165 is a greater chance that any benefits fall preferentially on the strain that invests in manipulation  
166 (Box 3). Potential benefits of such manipulation include increased nutrient supply and decreased  
167 inhibition by attenuating host immune responses. However, to explain effects on behaviour, any  
168 manipulation must also have side effects on the host central nervous system. One candidate  
169 route is through local changes to the enteric neurobiology of the host, which may then influence  
170 host behaviour through communication between the enteric and central nervous systems<sup>58</sup>. Gut  
171 bacteria can modulate intestinal motility<sup>59</sup> through metabolites – including short-chain fatty acids  
172 and bile acids – that affect serotonin synthesis in the host<sup>60-63</sup>, and gut motility can in turn  
173 influence the competitive ability of certain species<sup>64</sup>. However, to our knowledge, there is not yet  
174 evidence that bacteria benefit from modifying gut motor function in a manner that would  
175 support the evolution of manipulation. Nevertheless, effects on the host enteric nervous system  
176 such as serotonergic signalling have the potential to modulate host mood and behaviour through  
177 the gut-brain connection.

178 Symbiotic microorganisms may also have local effects on host immune responses<sup>65,66</sup>,  
179 including reducing the host inflammatory response<sup>67,68</sup>. However, it is not clear whether these  
180 effects represent local manipulation by the microorganisms or arise purely as a function of  
181 natural selection on the host to discriminate between different microbial phenotypes<sup>69</sup>.  
182 Nevertheless, the immune and nervous systems are extensively connected<sup>70-72</sup>, not only  
183 mechanistically but also anatomically<sup>73</sup>, such that any microbial effects on the immune system  
184 may elicit behavioural changes as a side effect (without any natural selection on symbionts to  
185 manipulate behaviour). There is the possibility, therefore, that many of the effects described in  
186 studies of the gut-brain axis may actually reflect an immune response. Consistent with this,  
187 recent findings show that colonising mice with the faecal microbiota from patients with irritable  
188 bowel syndrome can drive anxiety-like behaviour, but only when mice also exhibit immune  
189 activation<sup>74</sup>.

190 The evolutionary basis for local manipulation by symbionts is on more solid ground than  
191 the global manipulation of host phenotypes. As such, some effects of the microbiota on host  
192 behaviour may be a side effect of local manipulation. However, even at the local scale, more data  
193 are needed to convincingly demonstrate the evolution of symbiont manipulation of the host, and  
194 distinguish it from the evolution of host adaptations that serve to detect and respond to  
195 particular strains and species in the gut<sup>75</sup>.

196

### 197 **By-products of microbial metabolism**

198 If the microbial compounds that affect host behaviour do not arise for manipulation, why are  
199 they produced? The simplest explanation is that they are generated as part of the metabolism  
200 that helps the microorganism to grow and divide, as occurs with metabolic waste products. Short-  
201 chain fatty acids (for example, butyrate, propionate, acetate) are key waste products made by  
202 gut bacteria, which can influence gut motility<sup>60-62</sup>, modulate host immune responses<sup>76</sup> and have  
203 substantial neuromodulatory effects, possibly because they function as histone deacetylase  
204 inhibitors<sup>77</sup>. Polysaccharide A, a component of the bacterial capsule, can also affect gut motility<sup>78</sup>  
205 and host immune responses<sup>79</sup>. In addition, microbial compounds may affect the brain. Butyrate  
206 helps maintain the integrity of the blood-brain barrier<sup>80</sup>, which typically functions to separate the  
207 neuroactive agents of the brain and periphery<sup>81</sup>. Furthermore, acetate produced in the colon can  
208 cross the blood-brain barrier and directly enter the brain<sup>82</sup>.

209 Compounds that function as host neurotransmitters (Fig. 2) are particularly relevant.  
210 *Lactobacillus* and *Bifidobacterium* species from the human intestine are prolific producers of  $\gamma$ -  
211 aminobutyric acid (GABA) in culture<sup>83</sup>. Moreover, expressing a *Bifidobacterium dentium* gene for  
212 GABA production in the mouse gut using transformed *B. breve* can modulate indicators of visceral  
213 pain<sup>84</sup>. A GABA uptake system has also been reported in *Pseudomonas fluorescens*, a plant-  
214 associated bacterium<sup>85</sup>, and more recently a bacterial species from the human gut microbiome,  
215 *Evtepia gabavorous*, has been shown to require GABA as a growth factor<sup>86</sup>. Uptake of a  
216 neurotransmitter provides another potential route to influence host behaviour. In addition,  
217 bacteria seem to have an important role in activating precursors of dopamine and noradrenaline  
218 in the gut<sup>87</sup>, and some species also synthesise serotonin, acetylcholine and histamine<sup>88</sup>. The  
219 production of these molecules raises the possibility that microorganism-derived  
220 neurotransmitters can bind directly to host receptors<sup>88</sup>.

221 Does the production of neurotransmitters identify symbionts that have evolved to  
222 manipulate the mammalian brain? This is far from clear. Firstly, neurotransmitters may not be  
223 produced at meaningful levels by bacteria in the gut, as studies describing bacterial  
224 neurotransmitter production are largely performed *in vitro*<sup>89</sup>. Secondly, it remains unknown  
225 whether lumen-produced neurotransmitters (or their precursors) can strongly influence the  
226 brain (Fig. 2). Moreover, even if microorganism-derived neurotransmitters can affect the brain,  
227 their production may well be explained by another bacterial function, rather than host

228 manipulation. Although these compounds are known as 'neurotransmitters' in animals, they are  
229 produced not only by bacteria but also fungi and plants<sup>90</sup>. Indeed, their use in multicellular  
230 species may even be explained by horizontal gene transfer events from bacteria<sup>91</sup>. Most  
231 importantly, bacteria isolated from the environment can also produce neurotransmitters<sup>90,92</sup>,  
232 which suggests that these compounds have a role in bacterial biology outside of the host. The  
233 function of these compounds in free-living bacteria, therefore, is an important open question; as  
234 is whether these functions translate to symbiotic species. Initial work in this area suggests  
235 functions in both core metabolism (for example, breakdown of amino acids) and signalling  
236 between cells<sup>90</sup>.

237

### 238 **The evolution of host dependence**

239 Our focus has been on the microbiota and how microbial evolution can lead to effects on the  
240 host. However, it is also important that we consider how host evolution can influence the  
241 microbiota-gut-brain axis. Here there are several non-mutually exclusive routes for natural  
242 selection on hosts to affect or forge links from the microbiome to behaviour. The simplest stems  
243 from the possibility that a host behaves differently when certain microorganisms are lacking  
244 simply because its physiology is compromised. Such effects can have multiple causes, but  
245 evolutionarily they are expected from what is often called 'evolved dependence'<sup>93,94</sup>. When a  
246 host evolves alongside a symbiont, even a harmful one, there is the potential for it to come to  
247 rely on that symbiont for certain functions. For example, the wasp *Asobara tabida* has evolved  
248 to depend on the bacterial endosymbiont *Wolbachia* species for normal oocyte development,  
249 even though this bacterium is commonly a parasite of insects<sup>95</sup>.

250 Evolved dependence may affect both the nervous system and immune system, such that  
251 removing a particular microorganism creates a maladaptive physiological state that translates to  
252 behavioural effects. This can then lead to specific microorganisms having specific effects on host  
253 behaviour without any natural selection on the microorganisms to influence host physiology.  
254 Given the apparent functional redundancy of the gut microbiota<sup>96</sup>, multiple phylogenetically-  
255 diverse symbionts may complement any host dependence. Therefore, it may be more the loss of  
256 the microbial trait, rather than specific microbial species, that leads to any impairment in host  
257 behaviour. More generally, evolved dependence may explain why an altered gut microbiome  
258 composition (such as in the case of germ-free or antibiotic-treated animals) is associated with  
259 behavioural changes<sup>97-100</sup>. If we have evolved to depend on microbiota to modulate our own  
260 neurochemistry, then we may expect their absence to influence brain function.

261 There is also the potential for evolved dependence through the evolution of the immune  
262 system. The long evolutionary association with symbiotic microbiota has provided many  
263 opportunities for immune regulation to evolve dependencies on bacterial phenotypes. Broadly  
264 consistent with this, gut microbiota affect various aspects of the host immune  
265 response<sup>76,79,101,102</sup>. For example, microbial metabolites influence the differentiation and

266 functioning of immune cells<sup>103,104</sup> and can have anti-inflammatory effects<sup>105</sup>. Evolved  
267 dependence has also been linked to the hygiene hypothesis, which posits a causal association  
268 between improved hygiene and the rise in autoimmune conditions<sup>94,106</sup>. This is based on the idea  
269 that an absence of symbiotic microorganisms or parasites leads to immune dysregulation. Most  
270 relevant here is the suggestion that the hygiene hypothesis may be linked to mental health  
271 through neuroimmune connections<sup>107</sup>. Thus, immune processes may underpin many of the  
272 effects of the microbiome on the brain. Indeed, the bacterial genera *Lactobacillus* and  
273 *Bifidobacterium* that are commonly associated with behavioural changes are also known for their  
274 immunomodulatory properties<sup>108,109</sup>. Although the study of the microbiota-gut-brain axis has not  
275 explicitly considered evolved dependence, we believe it may prove fundamental to how the  
276 microbiome affects the brain.

277 Evolved dependence rests upon the idea that host physiology may come to depend upon  
278 symbionts for normal functioning. Host evolution can also generate new functions through the  
279 microbiota, and these may again affect behaviour. The gut microbiota contains much information  
280 of value to a host. When the microbiota inform on nutritional state, natural selection on the host  
281 may link the state of the microbiota to host appetite, feeding and foraging behaviour<sup>24,110</sup>. For  
282 example, short-chain fatty acids produced by microbial fermentation are implicated in satiety  
283 regulation<sup>111</sup>. Another potential evolved response is sickness behaviour resulting from the spread  
284 of a pathogen within the gut (Box 1). Furthermore, when particular symbionts provide valuable  
285 information or perform a useful function, even if just through a by-product of microbial  
286 metabolism, a host may evolve mechanisms to favour these bacterial species and thereby  
287 reinforce their effects. There are many potential routes to such host control of the microbiota by  
288 compounds secreted from the host epithelium, including both specific nutrients and  
289 antimicrobials<sup>75</sup>. Thus, hosts are expected to evolve to depend upon, monitor and regulate their  
290 microbiota. This evolution may readily forge and modulate links between the microbiota and host  
291 behaviour.

292

## 293 **Outlook**

294 There is growing evidence that the mammalian gut microbiota have effects on the brain and  
295 behaviour, raising the hypothesis that our microbiota have evolved to manipulate us<sup>23–28</sup>.  
296 However, taking stock of both data and evolutionary theory casts serious doubt on this  
297 hypothesis. The theory predicts that manipulation is most likely when the manipulative trait has  
298 low cost and high benefit for the manipulating bacteria, and critically when there is limited  
299 competition from non-manipulating strains (Table 1). This last condition does not seem easily  
300 satisfied in the diverse microbial ecosystem of the gut. We should not then assume that our  
301 microbiota are our puppeteers. Instead, the behavioural effects of the microbiota appear better  
302 explained as a side effect of either local manipulation of the host environment or the microbial  
303 metabolism needed to grow and survive in the gut. Moreover, it is clear that hosts can evolve to



304 depend upon the microbiota, and use them to respond to nutritional and disease states, thereby  
305 cementing a link from symbionts to host physiology.

306 Our perspective has implications for both understanding and manipulating how the  
307 microbiota affect behaviour. We predict that microbial compounds that influence host  
308 physiology, such as neurotransmitters, typically evolve either because of their local impacts on  
309 host physiology (local manipulation), or as a by-product of natural selection on microorganisms  
310 to grow and compete within the microbiota (metabolic by-product, Fig. 1). Local effects on mucus  
311 production<sup>41,112</sup>, the inflammatory response<sup>56</sup> and gut motility<sup>64</sup> all have the potential to  
312 influence microbial strains differently, in ways important for evolutionary success. However, the  
313 clearest evidence for local manipulation currently comes from acute pathogens such as  
314 *Salmonella* strains, rather than from beneficial or commensal symbionts. The demonstration of  
315 local manipulation by symbionts requires more than simply showing effects of a microorganism  
316 on host physiology. Critically, local manipulation is also predicted to increase the competitive  
317 ability of the manipulating strain in the gut, in contrast to behavioural manipulation which is not  
318 expected to improve local competitive ability (Table 1). For symbionts that spend a long time in  
319 the host, this predicts that a locally manipulating strain will outcompete an otherwise isogenic  
320 strain lacking the manipulative trait. Moreover, this competitive benefit must arise from effects  
321 on host physiology.

322 Arguably, the simplest explanation for microbial traits that influence host behaviour is  
323 that they are a by-product of the way that microorganisms grow and compete in the gut (Fig. 1).  
324 Similar to local manipulation, this predicts that the production of a host-affecting compound will  
325 provide a competitive advantage to the strain that carries it. However, in contrast to local  
326 manipulation, the advantage will occur independently of the effects on host physiology. This  
327 implies that any advantage can also be observed experimentally *ex vivo* (to the extent that the  
328 experiment can capture the growth conditions in the host). What kinds of molecule help a  
329 microorganism to compete but might also affect host physiology? There are many candidates.  
330 We have focussed our discussion on metabolic products such as short-chain fatty acids that are  
331 known to strongly influence the physiology of host cells<sup>60–62,76</sup>. However, bacteria produce vast  
332 numbers of compounds whose effects on gut physiology are currently unknown<sup>113</sup>.

333 Metabolic waste products are just one source of compounds released by bacteria. To  
334 compete in any community, microorganisms produce a wide variety of compounds that influence  
335 the survival and division of other cells (Box 3). These include enzymes that break down complex  
336 molecules, bio-surfactants, siderophores that scavenge iron, diverse toxins that inhibit other  
337 microorganisms, extracellular polymeric substances including carbohydrates and DNA, molecules  
338 that function as electron acceptors, and molecules that serve in cell-cell communication (quorum  
339 sensing)<sup>114,115</sup>. Such compounds are also potential candidates for influencing host physiology  
340 because they can be released in large quantities and, moreover, have often evolved because of  
341 their physiological effects on other cells. For example, iron is a key currency for both host and

342 microbial cells<sup>116</sup> and bacterial siderophores that scavenge iron are known to affect epithelial cell  
343 physiology<sup>117</sup>.

344 A focus on ecology also has implications for the goal of engineering the microbiota<sup>75</sup>. It  
345 has been suggested that probiotic strains might be used to improve mental health outcomes<sup>19</sup>. A  
346 major challenge with many probiotics is getting a strain to establish itself in a new  
347 community<sup>75,118</sup>. However, a focus on naturally-occurring human symbionts should circumvent  
348 this challenge. If we are correct that host-affecting traits are accompanied by a competitive  
349 advantage, probiotic strategies should be viable in the sense that the strains can compete and  
350 establish themselves in communities. Another limitation of probiotic strategies is the tendency  
351 to seek a single strain to provide a given benefit. In reality, the benefits that the microbiota  
352 provide to the host, such as protection against pathogens, can arise from the interactions of  
353 multiple species within a community<sup>119</sup>. If this is also true for behavioural effects, then we will  
354 need to embrace the full ecological complexity of the microbiota in order to understand the gut-  
355 brain axis.

356

## 357 References

- 358 1. Mayer, E. A. Gut feelings: the emerging biology of gut-brain communication. *Nat. Rev. Neurosci.*  
359 **12**, 453–466 (2011).
- 360 2. Forsythe, P., Bienenstock, J. & Kunze, W. A. in *Microbial Endocrinology: The Microbiota-Gut-Brain*  
361 *Axis in Health and Disease* (eds Lyte, M. & Cryan, J. F.) 115–133 (Springer, New York, 2014).
- 362 3. Fung, T. C., Olson, C. A. & Hsiao, E. Y. Interactions between the microbiota, immune and nervous  
363 systems in health and disease. *Nat. Neurosci.* **20**, 145–155 (2017).
- 364 4. Neuman, H., Debelius, J. W., Knight, R. & Koren, O. Microbial endocrinology: the interplay between  
365 the microbiota and the endocrine system. *FEMS Microbiol. Rev.* **39**, 509–521 (2015).
- 366 5. Lyte, M. Microbial endocrinology in the microbiome-gut-brain axis: how bacterial production and  
367 utilization of neurochemicals influence behavior. *PLoS Pathog.* **9**, e1003726 (2013).
- 368 6. Cryan, J. F. & Dinan, T. G. Mind-altering microorganisms: the impact of the gut microbiota on brain  
369 and behaviour. *Nat. Rev. Neurosci.* **13**, 701–712 (2012).
- 370 7. Foster, J. A. & McVey Neufeld, K.-A. Gut-brain axis: how the microbiome influences anxiety and  
371 depression. *Trends Neurosci.* **36**, 305–312 (2013).
- 372 8. Sarkar, A. *et al.* Psychobiotics and the manipulation of bacteria-gut-brain signals. *Trends Neurosci.*  
373 **39**, 763–781 (2016).
- 374 9. Sharon, G., Sampson, T. R., Geschwind, D. H. & Mazmanian, S. K. The central nervous system and  
375 the gut microbiome. *Cell* **167**, 915–932 (2016).
- 376 10. Rhee, S. H., Pothoulakis, C. & Mayer, E. A. Principles and clinical implications of the brain-gut-  
377 enteric microbiota axis. *Nat. Rev. Gastroenterol. Hepatol.* **6**, 306–314 (2009).
- 378 11. Bercik, P. *et al.* The intestinal microbiota affect central levels of brain-derived neurotrophic factor  
379 and behavior in mice. *Gastroenterology* **141**, 599–609 (2011).
- 380 12. Desbonnet, L. *et al.* Effects of the probiotic *Bifidobacterium infantis* in the maternal separation  
381 model of depression. *Neuroscience* **170**, 1179–1188 (2010).
- 382 13. Bercik, P. *et al.* The anxiolytic effect of *Bifidobacterium longum* NCC3001 involves vagal pathways  
383 for gut-brain communication. *Neurogastroenterol. Motil.* **23**, 1132–1139 (2011).
- 384 14. Bravo, J. A. *et al.* Ingestion of *Lactobacillus* strain regulates emotional behavior and central GABA

- 385 receptor expression in a mouse via the vagus nerve. *Proc. Natl. Acad. Sci. USA* **108**, 16050–16055  
386 (2011).
- 387 15. Messaoudi, M. *et al.* Assessment of psychotropic-like properties of a probiotic formulation  
388 (*Lactobacillus helveticus* R0052 and *Bifidobacterium longum* R0175) in rats and human subjects.  
389 *Br. J. Nutr.* **105**, 755–764 (2011).
- 390 16. Savignac, H. M., Kiely, B., Dinan, T. G. & Cryan, J. F. *Bifidobacteria* exert strain-specific effects on  
391 stress-related behavior and physiology in BALB/c mice. *Neurogastroenterol. Motil.* **26**, 1615–1627  
392 (2014).
- 393 17. Davis, D. J. *et al.* *Lactobacillus plantarum* attenuates anxiety-related behavior and protects against  
394 stress-induced dysbiosis in adult zebrafish. *Sci. Rep.* **6**, 33726 (2016).
- 395 18. Pinto-Sanchez, M. I. *et al.* Probiotic *Bifidobacterium longum* NCC3001 reduces depression scores  
396 and alters brain activity: a pilot study in patients with irritable bowel syndrome. *Gastroenterology*  
397 **153**, 448–459 (2017).
- 398 19. Wallace, C. J. K. & Milev, R. The effects of probiotics on depressive symptoms in humans: a  
399 systematic review. *Ann. Gen. Psychiatry* **16** (2017).
- 400 20. Bharwani, A., Mian, M. F., Surette, M. G., Bienenstock, J. & Forsythe, P. Oral treatment with  
401 *Lactobacillus rhamnosus* attenuates behavioural deficits and immune changes in chronic social  
402 stress. *BMC Med.* **15**, 7 (2017).
- 403 21. Buffington, S. A. *et al.* Microbial reconstitution reverses maternal diet-induced social and synaptic  
404 deficits in offspring. *Cell* **165**, 1762–1775 (2016).
- 405 22. Hsiao, E. Y. *et al.* Microbiota modulate behavioral and physiological abnormalities associated with  
406 neurodevelopmental disorders. *Cell* **155**, 1451–1463 (2013).
- 407 23. Alcock, J., Maley, C. C. & Aktipis, C. A. Is eating behavior manipulated by the gastrointestinal  
408 microbiota? Evolutionary pressures and potential mechanisms. *BioEssays* **36**, 940–949 (2014).
- 409 24. Wong, A. C.-N. *et al.* Behavioral microbiomics: a multi-dimensional approach to microbial influence  
410 on behavior. *Front. Microbiol.* **6**, 1359 (2015).
- 411 25. Stilling, R. M., Dinan, T. G. & Cryan, J. F. The brain's Geppetto—microbes as puppeteers of neural  
412 function and behaviour? *J. Neurovirol.* **22**, 14–21 (2016).
- 413 26. Yuval, B. Symbiosis: gut bacteria manipulate host behaviour. *Curr. Biol.* **27**, R746–R747 (2017).
- 414 27. Stilling, R. M., Bordenstein, S. R., Dinan, T. G. & Cryan, J. F. Friends with social benefits: host-  
415 microbe interactions as a driver of brain evolution and development? *Front. Cell. Infect. Microbiol.*  
416 **4**, 147 (2014).
- 417 28. Lewin-Epstein, O., Aharonov, R. & Hadany, L. Microbes can help explain the evolution of host  
418 altruism. *Nat. Commun.* **8**, 14040 (2017).
- 419 29. Brown, S. P. Do all parasites manipulate their hosts? *Behav. Processes* **68**, 237–240 (2005).
- 420 30. Thomas, F., Adamo, S. A. & Moore, J. Parasitic manipulation: where are we and where should we  
421 go? *Behav. Processes* **68**, 185–199 (2005).
- 422 31. Adamo, S. A. Modulating the modulators: parasites, neuromodulators and host behavioral change.  
423 *Brain Behav. Evol.* **60**, 370–377 (2002).
- 424 32. Perrot-Minnot, M.-J. & Cézilly, F. Investigating candidate neuromodulatory systems underlying  
425 parasitic manipulation: concepts, limitations and prospects. *J. Exp. Biol.* **216**, 134–141 (2013).
- 426 33. Andersen, S. B. *et al.* The life of a dead ant: the expression of an adaptive extended phenotype.  
427 *Am. Nat.* **174**, 424–433 (2009).
- 428 34. Hughes, D. P. *et al.* Behavioral mechanisms and morphological symptoms of zombie ants dying  
429 from fungal infection. *BMC Ecol.* **11**, 13 (2011).
- 430 35. Klein, S. L. Parasite manipulation of the proximate mechanisms that mediate social behavior in  
431 vertebrates. *Physiol. Behav.* **79**, 441–449 (2003).
- 432 36. Berdoy, M., Webster, J. P. & Macdonald, D. W. Fatal attraction in rats infected with *Toxoplasma*

- 433 *gondii*. *Proc. R. Soc. Lond. B Biol. Sci.* **267**, 1591–1594 (2000).
- 434 37. Vyas, A., Kim, S.-K., Giacomini, N., Boothroyd, J. C. & Sapolsky, R. M. Behavioral changes induced  
435 by *Toxoplasma* infection of rodents are highly specific to aversion of cat odors. *Proc. Natl. Acad.*  
436 *Sci. USA* **104**, 6442–6447 (2007).
- 437 38. Poulin, R. ‘Adaptive’ changes in the behaviour of parasitized animals: a critical review. *Int. J.*  
438 *Parasitol.* **25**, 1371–1383 (1995).
- 439 39. Brown, S. P. Cooperation and conflict in host-manipulating parasites. *Proc. R. Soc. Lond. B Biol. Sci.*  
440 **266**, 1899–1904 (1999).
- 441 40. Vickery, W. L. & Poulin, R. The evolution of host manipulation by parasites: a game theory analysis.  
442 *Evol. Ecol.* **24**, 773–788 (2010).
- 443 41. Schluter, J. & Foster, K. R. The evolution of mutualism in gut microbiota via host epithelial selection.  
444 *PLoS Biol.* **10**, e1001424 (2012).
- 445 42. Xavier, J. B., Kim, W. & Foster, K. R. A molecular mechanism that stabilizes cooperative secretions  
446 in *Pseudomonas aeruginosa*. *Mol. Microbiol.* **79**, 166–179 (2011).
- 447 43. Adamo, S. A. Parasites: evolution’s neurobiologists. *J. Exp. Biol.* **216**, 3–10 (2013).
- 448 44. Bäckhed, F., Ley, R. E., Sonnenburg, J. L., Peterson, D. A. & Gordon, J. I. Host-bacterial mutualism  
449 in the human intestine. *Science* **307**, 1915–1920 (2005).
- 450 45. Qin, J. *et al.* A human gut microbial gene catalogue established by metagenomic sequencing.  
451 *Nature* **464**, 59–65 (2010).
- 452 46. Coyte, K. Z., Schluter, J. & Foster, K. R. The ecology of the microbiome: networks, competition, and  
453 stability. *Science* **350**, 663–666 (2015).
- 454 47. Stein, R. R. *et al.* Ecological modeling from time-series inference: insight into dynamics and stability  
455 of intestinal microbiota. *PLoS Comput. Biol.* **9**, e1003388 (2013).
- 456 48. Marino, S., Baxter, N. T., Huffnagle, G. B., Petrosino, J. F. & Schloss, P. D. Mathematical modeling  
457 of primary succession of murine intestinal microbiota. *Proc. Natl. Acad. Sci.* **111**, 439–444 (2014).
- 458 49. Kommineni, S. *et al.* Bacteriocin production augments niche competition by enterococci in the  
459 mammalian gastrointestinal tract. *Nature* **526**, 719–722 (2015).
- 460 50. Chatzidaki-Livanis, M., Geva-Zatorsky, N. & Comstock, L. E. *Bacteroides fragilis* type VI secretion  
461 systems use novel effector and immunity proteins to antagonize human gut Bacteroidales species.  
462 *Proc. Natl. Acad. Sci. USA* **113**, 3627–3632 (2016).
- 463 51. Wexler, A. G. *et al.* Human symbionts inject and neutralize antibacterial toxins to persist in the gut.  
464 *Proc. Natl. Acad. Sci. USA* **113**, 3639–3644 (2016).
- 465 52. Rao, S. *et al.* Pathogen-mediated inhibition of anorexia promotes host survival and transmission.  
466 *Cell* **168**, 503–516 (2017).
- 467 53. Murray, M. J. & Murray, A. B. Anorexia of infection of host defense as a mechanism. *Am. J. Clin.*  
468 *Nutr.* **32**, 593–596 (1979).
- 469 54. Wickham, M. E., Brown, N. F., Provias, J., Finlay, B. B. & Coombes, B. K. Oral infection of mice  
470 with *Salmonella enterica* serovar Typhimurium causes meningitis and infection of the brain. *BMC*  
471 *Infect. Dis.* **7**, 65 (2007).
- 472 55. David, L. A. *et al.* Host lifestyle affects human microbiota on daily timescales. *Genome Biol.* **15**, R89  
473 (2014).
- 474 56. Rivera-Chávez, F. & Bäumlér, A. J. The pyromaniac inside you: *Salmonella* metabolism in the host  
475 gut. *Annu. Rev. Microbiol.* **69**, 31–48 (2015).
- 476 57. Balmer, O. & Tanner, M. Prevalence and implications of multiple-strain infections. *Lancet Infect.*  
477 *Dis.* **11**, 868–878 (2011).
- 478 58. Rao, M. & Gershon, M. D. The bowel and beyond: the enteric nervous system in neurological  
479 disorders. *Nat. Rev. Gastroenterol. Hepatol.* **13**, 517–528 (2016).
- 480 59. Quigley, E. M. M. Microflora modulation of motility. *J. Neurogastroenterol. Motil.* **17**, 140–147

- 481 (2011).
- 482 60. Fukumoto, S. *et al.* Short-chain fatty acids stimulate colonic transit via intraluminal 5-HT release in  
483 rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **284**, R1269–R1276 (2003).
- 484 61. Reigstad, C. S. *et al.* Gut microbes promote colonic serotonin production through an effect of short-  
485 chain fatty acids on enterochromaffin cells. *FASEB J.* **29**, 1395–1403 (2015).
- 486 62. Yano, J. M. *et al.* Indigenous bacteria from the gut microbiota regulate host serotonin biosynthesis.  
487 *Cell* **161**, 264–276 (2015).
- 488 63. Dey, N. *et al.* Regulators of gut motility revealed by a gnotobiotic model of diet-microbiome  
489 interactions related to travel. *Cell* **163**, 95–107 (2015).
- 490 64. Wiles, T. J. *et al.* Host gut motility promotes competitive exclusion within a model intestinal  
491 microbiota. *PLoS Biol.* **14**, e1002517 (2016).
- 492 65. Sansonetti, P. J. & Di Santo, J. P. Debugging how bacteria manipulate the immune response.  
493 *Immunity* **26**, 149–161 (2007).
- 494 66. Ayres, J. S. Cooperative microbial tolerance behaviors in host-microbiota mutualism. *Cell* **165**,  
495 1323–1331 (2016).
- 496 67. Neish, A. S. *et al.* Prokaryotic regulation of epithelial responses by inhibition of I $\kappa$ B- $\alpha$  ubiquitination.  
497 *Science* **289**, 1560–1563 (2000).
- 498 68. Kelly, D. *et al.* Commensal anaerobic gut bacteria attenuate inflammation by regulating nuclear-  
499 cytoplasmic shuttling of PPAR- $\gamma$  and RelA. *Nat. Immunol.* **5**, 104–112 (2004).
- 500 69. Hooper, L. V. Do symbiotic bacteria subvert host immunity? *Nat. Rev. Microbiol.* **7**, 367–374 (2009).
- 501 70. Steinman, L. Elaborate interactions between the immune and nervous systems. *Nat. Immunol.* **5**,  
502 575–581 (2004).
- 503 71. Baganz, N. L. & Blakely, R. D. A dialogue between the immune system and brain, spoken in the  
504 language of serotonin. *ACS Chem. Neurosci.* **4**, 48–63 (2013).
- 505 72. Wohleb, E. S., Franklin, T., Iwata, M. & Duman, R. S. Integrating neuroimmune systems in the  
506 neurobiology of depression. *Nat. Rev. Neurosci.* **17**, 497–511 (2016).
- 507 73. Louveau, A. *et al.* Structural and functional features of central nervous system lymphatic vessels.  
508 *Nature* **523**, 337–341 (2015).
- 509 74. De Palma, G. *et al.* Transplantation of fecal microbiota from patients with irritable bowel syndrome  
510 alters gut function and behavior in recipient mice. *Sci. Transl. Med.* **9**, eaaf6397 (2017).
- 511 75. Foster, K. R., Schluter, J., Coyte, K. Z. & Rakoff-Nahoum, S. The evolution of the host microbiome  
512 as an ecosystem on a leash. *Nature* **548**, 43–51 (2017).
- 513 76. Rooks, M. G. & Garrett, W. S. Gut microbiota, metabolites and host immunity. *Nat. Rev. Immunol.*  
514 **16**, 341–352 (2016).
- 515 77. Koh, A., De Vadder, F., Kovatcheva-Datchary, P. & Bäckhed, F. From dietary fiber to host physiology:  
516 short-chain fatty acids as key bacterial metabolites. *Cell* **165**, 1332–1345 (2016).
- 517 78. Mao, Y.-K. *et al.* *Bacteroides fragilis* polysaccharide A is necessary and sufficient for acute activation  
518 of intestinal sensory neurons. *Nat. Commun.* **4**, 1465 (2013).
- 519 79. Mazmanian, S. K. & Kasper, D. L. The love–hate relationship between bacterial polysaccharides and  
520 the host immune system. *Nat. Rev. Immunol.* **6**, 849–858 (2006).
- 521 80. Braniste, V. *et al.* The gut microbiota influences blood-brain barrier permeability in mice. *Sci.*  
522 *Transl. Med.* **6**, 263ra158 (2014).
- 523 81. Abbott, N. J., Rönnbäck, L. & Hansson, E. Astrocyte-endothelial interactions at the blood-brain  
524 barrier. *Nat. Rev. Neurosci.* **7**, 41–53 (2006).
- 525 82. Frost, G. *et al.* The short-chain fatty acid acetate reduces appetite via a central homeostatic  
526 mechanism. *Nat. Commun.* **5**, 3611 (2014).
- 527 83. Barrett, E., Ross, R. P., O’Toole, P. W., Fitzgerald, G. F. & Stanton, C.  $\gamma$ -Aminobutyric acid production  
528 by culturable bacteria from the human intestine. *J. Appl. Microbiol.* **113**, 411–417 (2012).

- 529 84. Pokusaeva, K. *et al.* GABA-producing *Bifidobacterium dentium* modulates visceral sensitivity in the  
530 intestine. *Neurogastroenterol. Motil.* **29**, e12904 (2017).
- 531 85. Guthrie, G. D. & Nicholson-Guthrie, C. S.  $\gamma$ -Aminobutyric acid uptake by a bacterial system with  
532 neurotransmitter binding characteristics. *Proc. Natl. Acad. Sci. USA* **86**, 7378–7381 (1989).
- 533 86. Strandwitz, P. *et al.* GABA modulating bacteria of the human gut microbiome at *ASM Microbe*  
534 *Conference* (American Society for Microbiology, 2016).
- 535 87. Asano, Y. *et al.* Critical role of gut microbiota in the production of biologically active, free  
536 catecholamines in the gut lumen of mice. *Am. J. Physiol. Gastrointest. Liver Physiol.* **303**, G1288–  
537 G1295 (2012).
- 538 88. Wall, R. *et al.* in *Microbial Endocrinology: The Microbiota-Gut-Brain Axis in Health and Disease* (eds  
539 Lyte, M. & Cryan, J. F.) 221–239 (Springer, New York, 2014).
- 540 89. Sampson, T. R. & Mazmanian, S. K. Control of brain development, function, and behavior by the  
541 microbiome. *Cell Host Microbe* **17**, 565–576 (2015).
- 542 90. Roshchina, V. V. in *Microbial Endocrinology: Interkingdom Signaling in Infectious Disease and*  
543 *Health* (eds Lyte, M. & Freestone, P. P. E.) 17–52 (Springer, New York, 2010).
- 544 91. Iyer, L. M., Aravind, L., Coon, S. L., Klein, D. C. & Koonin, E. V. Evolution of cell-cell signaling in  
545 animals: did late horizontal gene transfer from bacteria have a role? *Trends Genet.* **20**, 292–299  
546 (2004).
- 547 92. Mountfort, D. O. & Pybus, V. Regulatory influences on the production of gamma-aminobutyric acid  
548 by a marine pseudomonad. *Appl. Environ. Microbiol.* **58**, 237–242 (1992).
- 549 93. de Mazancourt, C., Loreau, M. & Dieckmann, U. Understanding mutualism when there is  
550 adaptation to the partner. *J. Ecol.* **93**, 305–314 (2005).
- 551 94. Weinersmith, K. L. & Earley, R. L. Better with your parasites? Lessons for behavioural ecology from  
552 evolved dependence and conditionally helpful parasites. *Anim. Behav.* **118**, 123–133 (2016).
- 553 95. Pannebakker, B. A., Loppin, B., Elemans, C. P. H., Humblot, L. & Vavre, F. Parasitic inhibition of cell  
554 death facilitates symbiosis. *Proc. Natl. Acad. Sci. USA* **104**, 213–215 (2007).
- 555 96. Moya, A. & Ferrer, M. Functional redundancy-induced stability of gut microbiota subjected to  
556 disturbance. *Trends Microbiol.* **24**, 402–413 (2016).
- 557 97. Diaz Heijtz, R. *et al.* Normal gut microbiota modulates brain development and behavior. *Proc. Natl.*  
558 *Acad. Sci. USA* **108**, 3047–3052 (2011).
- 559 98. Desbonnet, L., Clarke, G., Shanahan, F., Dinan, T. G. & Cryan, J. F. Microbiota is essential for social  
560 development in the mouse. *Mol. Psychiatry* **19**, 146–148 (2014).
- 561 99. Desbonnet, L. *et al.* Gut microbiota depletion from early adolescence in mice: implications for brain  
562 and behaviour. *Brain. Behav. Immun.* **48**, 165–173 (2015).
- 563 100. Hoban, A. E. *et al.* Behavioural and neurochemical consequences of chronic gut microbiota  
564 depletion during adulthood in the rat. *Neuroscience* **339**, 463–477 (2016).
- 565 101. Round, J. L. & Mazmanian, S. K. The gut microbiota shapes intestinal immune responses during  
566 health and disease. *Nat. Rev. Immunol.* **9**, 313–323 (2009).
- 567 102. Hill, D. A. & Artis, D. Intestinal bacteria and the regulation of immune cell homeostasis. *Annu. Rev.*  
568 *Immunol* **28**, 623–667 (2010).
- 569 103. Furusawa, Y. *et al.* Commensal microbe-derived butyrate induces the differentiation of colonic  
570 regulatory T cells. *Nature* **504**, 446–450 (2013).
- 571 104. Smith, P. M. *et al.* The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell  
572 homeostasis. *Science* **341**, 569–573 (2013).
- 573 105. Brestoff, J. R. & Artis, D. Commensal bacteria at the interface of host metabolism and the immune  
574 system. *Nat. Immunol.* **14**, 676–684 (2013).
- 575 106. Strachan, D. P. Hay fever, hygiene, and household size. *Br. Med. J.* **299**, 1259–1260 (1989).
- 576 107. Rook, G. A. W. & Lowry, C. A. The hygiene hypothesis and psychiatric disorders. *Trends Immunol.*

- 577           **29**, 150–158 (2008).
- 578 108. Wells, J. M. Immunomodulatory mechanisms of lactobacilli. *Microb. Cell Fact.* **10** (Suppl. 1), S17  
579 (2011).
- 580 109. Fanning, S. *et al.* Bifidobacterial surface-exopolysaccharide facilitates commensal-host interaction  
581 through immune modulation and pathogen protection. *Proc. Natl. Acad. Sci. USA* **109**, 2108–2113  
582 (2012).
- 583 110. Fetissov, S. O. Role of the gut microbiota in host appetite control: bacterial growth to animal  
584 feeding behaviour. *Nat. Rev. Endocrinol.* **13**, 11–25 (2016).
- 585 111. Rosenbaum, M., Knight, R. & Leibel, R. L. The gut microbiota in human energy homeostasis and  
586 obesity. *Trends Endocrinol. Metab.* **26**, 493–501 (2015).
- 587 112. McLoughlin, K., Schluter, J., Rakoff-Nahoum, S., Smith, A. L. & Foster, K. R. Host selection of  
588 microbiota via differential adhesion. *Cell Host Microbe* **19**, 550–559 (2016).
- 589 113. Franzosa, E. A. *et al.* Sequencing and beyond: integrating molecular ‘omics’ for microbial  
590 community profiling. *Nat. Rev. Microbiol.* **13**, 360–372 (2015).
- 591 114. Mitri, S. & Foster, K. R. The genotypic view of social interactions in microbial communities. *Annu.*  
592 *Rev. Genet.* **47**, 247–273 (2013).
- 593 115. Nadell, C. D., Drescher, K. & Foster, K. R. Spatial structure, cooperation and competition in  
594 biofilms. *Nat. Rev. Microbiol.* **14**, 589–600 (2016).
- 595 116. Markel, T. A. *et al.* The struggle for iron: gastrointestinal microbes modulate the host immune  
596 response during infection. *J. Leukoc. Biol.* **81**, 393–400 (2007).
- 597 117. Choi, E.-Y. *et al.* Iron chelator triggers inflammatory signals in human intestinal epithelial cells:  
598 involvement of p38 and extracellular signal-regulated kinase signaling pathways. *J. Immunol.* **172**,  
599 7069–7077 (2004).
- 600 118. Weimer, P. J. Redundancy, resilience, and host specificity of the ruminal microbiota: implications  
601 for engineering improved ruminal fermentations. *Front. Microbiol.* **6**, 296 (2015).
- 602 119. Caballero, S. *et al.* Cooperating commensals restore colonization resistance to vancomycin-  
603 resistant *Enterococcus faecium*. *Cell Host Microbe* **21**, 592–602 (2017).
- 604 120. Thompson, J. N. *Interaction and Coevolution* (University of Chicago Press, Chicago, 1982).
- 605 121. Marchesi, J. R. & Ravel, J. The vocabulary of microbiome research: a proposal. *Microbiome* **3**, 31  
606 (2015).
- 607 122. Méthot, P.-O. & Alizon, S. What is a pathogen? Toward a process view of host-parasite interactions.  
608 *Virulence* **5**, 775–785 (2014).
- 609 123. May, G. & Nelson, P. Defensive mutualisms: do microbial interactions within hosts drive the  
610 evolution of defensive traits? *Funct. Ecol.* **28**, 356–363 (2014).
- 611 124. Hamilton, W. D. The genetical evolution of social behaviour I. *J. Theor. Biol.* **7**, 1–16 (1964).
- 612 125. Hamilton, W. D. The genetical evolution of social behaviour II. *J. Theor. Biol.* **7**, 17–52 (1964).
- 613 126. Bourke, A. F. G. *Principles of Social Evolution* (Oxford University Press, 2011).
- 614 127. Wilson, E. O. *Sociobiology: The New Synthesis* (Harvard University Press, 1975).
- 615 128. Sana, T. G., Lugo, K. A. & Monack, D. M. T6SS: the bacterial ‘fight club’ in the host gut. *PLoS Pathog.*  
616 **13**, e1006325 (2017).
- 617 129. Mitri, S. & Foster, K. R. Pleiotropy and the low cost of individual traits promote cooperation.  
618 *Evolution* **70**, 488–494 (2016).
- 619 130. LATERA, J., KEEP, R., BETZ, L. A. & GOLDSTEIN, G. W. in *Basic Neurochemistry: Molecular, Cellular and*  
620 *Medical Aspects* (eds Siegel, G. J., Agranoff, B. W., Albers, R. W., Fisher, S. K. & Uhler, M. D.) Ch. 32  
621 (Lippincott-Raven, Philadelphia, 1999).
- 622 131. Forsythe, P. & Kunze, W. A. Voices from within: gut microbes and the CNS. *Cell. Mol. Life Sci.* **70**,  
623 55–69 (2013).
- 624 132. Fernstrom, J. D. Role of precursor availability in control of monoamine biosynthesis in brain.

- 625 *Physiol. Rev.* **63**, 484–546 (1983).
- 626 133. Banks, W. A. Characteristics of compounds that cross the blood-brain barrier. *BMC Neurol.* **9**  
627 (Suppl. 1), S3 (2009).
- 628 134. O’Mahony, S. M., Clarke, G., Borre, Y. E., Dinan, T. G. & Cryan, J. F. Serotonin, tryptophan  
629 metabolism and the brain-gut-microbiome axis. *Behav. Brain Res.* **277**, 32–48 (2015).
- 630 135. Fernstrom, J. D. & Wurtman, R. J. Brain serotonin content: physiological dependence on plasma  
631 tryptophan levels. *Science* **173**, 149–152 (1971).
- 632 136. Biron, D. G. *et al.* Behavioural manipulation in a grasshopper harbouring hairworm: a proteomics  
633 approach. *Proc. R. Soc. Lond. B Biol. Sci.* **272**, 2117–2126 (2005).

634

635

### 636 **Acknowledgements**

637 We thank Sarah Knowles, Seth Rakoff-Nahoum, Elaine Hsiao, Joanne Webster and three  
638 anonymous reviewers for helpful comments on the manuscript.

639

### 640 **Author contributions**

641 K.V.-A.J. researched data for the article. Both authors substantially contributed to discussion of  
642 content, wrote the article and reviewed and edited the manuscript before submission.

643

### 644 **Competing interests statement**

645 The authors declare no financial or non-financial competing interests.

646

### 647 **Publisher's note**

648 Springer Nature remains neutral with regard to jurisdictional claims in published maps and  
649 institutional affiliations.

650

651 **Box 1. Examples of parasites affecting host behaviour.** The fungal parasite *Ophiocordyceps*  
652 *unilateralis* induces ants to reach a certain elevation in the canopy, where they then bite on  
653 vegetation with the so-called ‘death grip’<sup>33</sup> (figure, part a). This secures them at a position in the  
654 canopy that is favourable for fungal growth and the fungus then emerges from the base of the  
655 ant’s head to sporulate<sup>33</sup>. In vertebrates, parasite infections can change the social behaviour of  
656 hosts in ways that may promote parasite transmission<sup>35</sup>. For example, rabies virus infects  
657 mammals, including dogs and humans. The virus causes inflammation of the central nervous  
658 system and increased host aggression (figure, part b), which leads to biting and transmission<sup>35</sup>.  
659 The protozoan parasite *Toxoplasma gondii* infects birds and mammals and has been shown to



660 reduce the aversion of rodents to cat urine<sup>36,37</sup> (figure, part c). This puts the rodent at greater risk  
661 of predation and increases the chance of parasite transmission to feline hosts, which is necessary  
662 for the parasite to reproduce sexually<sup>36</sup>. Infection can cause sickness behaviour in hosts, including  
663 behaviours such as appetite loss<sup>53</sup> (figure, part d). The evolutionary basis for sickness behaviour  
664 is not always clear but one feature, loss of appetite, may have evolved to decrease nutrient  
665 supply to intestinal pathogens. Interestingly, there is evidence that *Salmonella enterica* serovar  
666 Typhimurium suppresses this appetite loss, which may represent manipulation of host feeding  
667 behaviour<sup>52</sup>.



## 680 **Box 2. The semantics of host-microbiota systems**

681 Diverse definitions abound in the study of host-microbiota systems, but the fields of ecology and  
682 evolution have a set of mostly-agreed definitions that can be applied consistently to avoid  
683 confusion. Here we outline these definitions, along with those of the microbiota and microbiome:

684 **Coevolution:** reciprocal evolutionary adaptations in different species in response to one  
685 another. If species A changes, species B changes in response and, critically, this feeds back and  
686 then species A changes again<sup>120</sup>.

688 **Commensalism:** interaction between species in which individuals on one side receive net fitness  
689 benefits whereas the other species are unaffected.

690 **Commensal:** party in a commensalism that receives benefit but has no net fitness effect on the  
691 other party.

692 **Competition:** interaction between species in which individuals on both sides suffer net fitness  
693 costs.

694 **Manipulation:** a manipulating symbiont alters the host phenotype in such a way as to improve  
695 the fitness of the symbiont, enabling its frequency within the population to increase. For  
696 example, symbiont fitness may be increased by increased transmission to new hosts or increased  
697 access to resources.

698 **Microbiome:** the community of microorganisms plus the environment. In host-associated  
699 microorganisms, this translates to the microbiota plus the host environment. This follows the  
700 proposed definition<sup>121</sup> and logically stems from the meaning of 'biome' as a major type of  
701 ecological community. Others limit the definition of the microbiome to the genomic material of  
702 the microbiota.

703 **Microbiota:** a community of microorganisms associated with a particular environment.

704 **Mutualism:** interaction between species in which all individuals receive net fitness benefits from  
705 the interaction.

706 **Parasitism:** interaction between species in which individuals on one side receive net fitness  
707 benefits whereas the other species experience net fitness costs. Parasites can be members of the  
708 microbiota with similar ecologies to commensal and mutualistic microorganisms.

709 **Symbiosis:** close ecological interaction between organisms (translated from the Greek meaning  
710 'living with'). Examples of symbiosis include mutualism, parasitism, commensalism and more.

711 **Symbiont:** member of a symbiosis that lives in or on the other member.

712 These definitions highlight that the mammalian microbiota are best described as symbionts,  
713 rather than the commonly used commensals because the former is silent on their potentially  
714 varied effects on the host. Indeed, one limitation of definitions based on fitness benefits is that  
715 a single symbiont may switch, for example, from mutualist to parasite under certain  
716 conditions<sup>122</sup>, making their classification challenging without a full knowledge of their effects<sup>123</sup>.

717

### 718 **Box 3. Social evolution, relatedness and host manipulation**

719 Our prediction that the microbiota rarely manipulate mammalian hosts originates from the field  
720 of social evolution<sup>124-127</sup>. Social evolutionists seek to understand the origin of traits in one  
721 organism that affect the survival and reproduction of other individuals. A classic example is the  
722 sterile, and sometimes suicidal, workers of insect societies. Such phenotypes, that harm the  
723 reproduction of the individual but benefit others, are known as altruism in evolutionary biology.  
724 Altruistic traits can evolve when there is genetic similarity between the carrier and the benefiting  
725 recipients<sup>124-126</sup>, because this means that an actor can increase the propagation of its alleles  
726 through the copies in a recipient. More specifically, the key determinant in social evolution is that

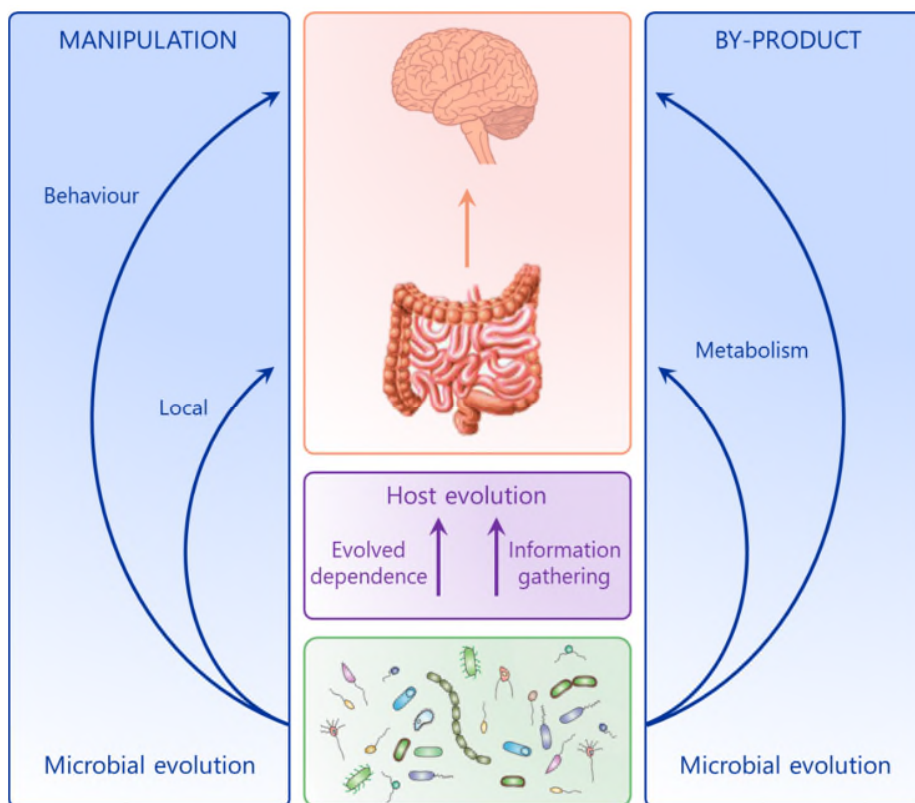
727 of 'relatedness', which captures the genetic similarity between individuals at the locus that drives  
728 the altruistic trait, relative to the population average. The main way to create relatedness is  
729 family life; the evolution of sterility in workers is explained by the fact that the queen in the  
730 colony is typically the mother of the workers. This means that the workers are raising siblings and  
731 are therefore able to pass on their genetic information, even though they do not themselves  
732 reproduce.

733 In microorganisms such as bacteria, relatedness emerges easily by binary fission, which  
734 can create a large group of a single genotype. At the scale of such groups, cooperative  
735 phenotypes in which several bacterial cells work together are extremely common, including the  
736 production of signalling molecules, enzymes to break down complex molecules and siderophores  
737 that scavenge iron<sup>115</sup>. However, beyond the scale of a clonal group, competition between  
738 genotypes (through both nutrient acquisition and the many toxins used by strains to kill others)  
739 is commonly predicted and observed<sup>114</sup>. The challenge to host manipulation then is that multiple  
740 competing strains can benefit, whereas only the strain that actually invests in manipulation will  
741 experience the cost, putting it at a disadvantage.

742 The problem of competition for manipulation was realised over fifteen years ago in a  
743 seminal social evolution paper, which predicted a positive relationship between relatedness  
744 within a group of parasites in a host and potential investment in host manipulation<sup>39</sup>. Although  
745 caution is required when applying relatedness measures to microbial communities, in which  
746 many strains and species may compete and share genes<sup>114</sup>, this prediction from the parasitology  
747 literature<sup>39</sup> remains relevant for the mammalian microbiota. Two sources of competition  
748 threaten to undermine a manipulating strain, one being strain diversity within its niche. If there  
749 are many different competing microorganisms, genetic relatedness will be very low, which  
750 disfavors any trait that costs a manipulating strain but benefits all others at the scale of the  
751 host<sup>41,75</sup>. If a manipulating strain can prevent immigration of other strains into its niche, the  
752 prospects for manipulation are improved. Such colonisation resistance is seen in the microbiota,  
753 and some species like *Bacteroides fragilis* often seem to occur as a single strain within a host<sup>128</sup>.  
754 However, even for such cases, a manipulating strain may be outcompeted by a second source of  
755 competitors: a mutant in the genetic background of the strain that lacks the manipulative trait.  
756 Low costs to manipulation and genetic constraints on the emergence of loss-of-function mutants  
757 may slow this process<sup>129</sup>. Nevertheless, the expectation is that a manipulative trait will be lost  
758 under long-term competition in the mammalian gut as any small growth cost associated with  
759 manipulation can drive the loss of a strain given the many microbial generations that commonly  
760 occur within the lifetime of a host<sup>41</sup>.

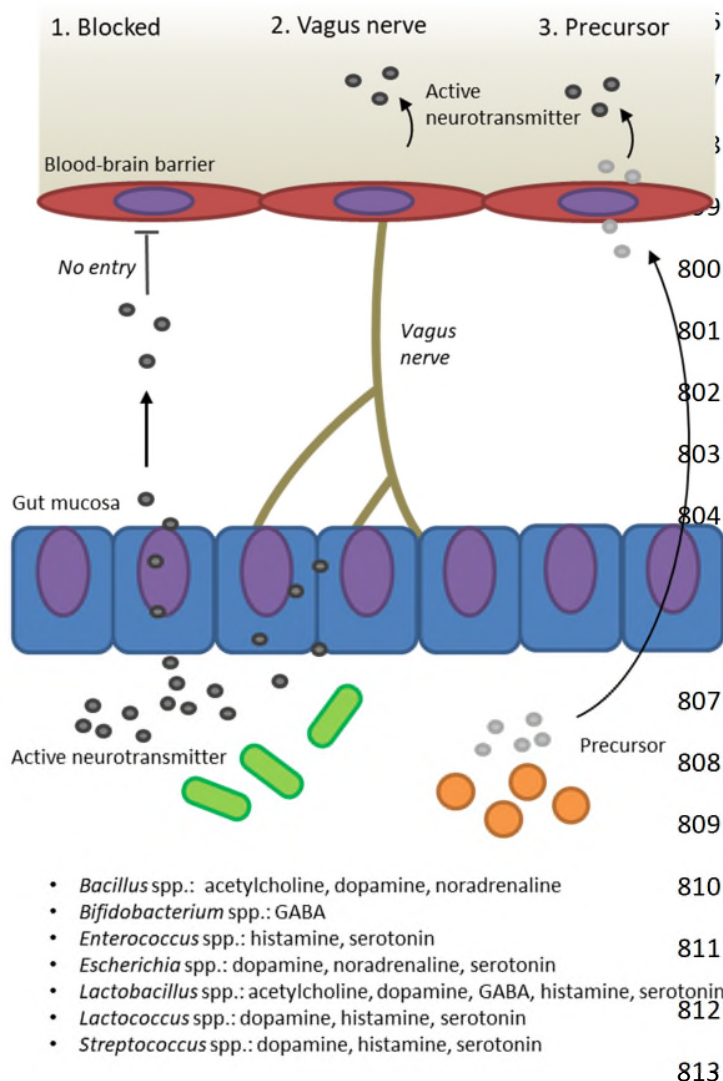
761

762



776

777 **Figure 1. Evolution of microbial effects on the brain.** Arrows denote the potential routes by  
 778 which microorganisms may influence host behaviour. Effects driven by natural selection on the  
 779 microbiota are shown in blue. The left-hand side captures microbial manipulation, in which case  
 780 the effects on the host increase microbial fitness. Here, the microbiota-gut-brain axis arises as an  
 781 evolutionary adaptation of microorganisms to influence either the gut environment (local  
 782 manipulation of host physiology) or host behaviour (global manipulation of the host). The right-  
 783 hand side depicts the evolution of microbial traits that affect the brain without the evolution of  
 784 manipulation. For example, the evolution of the metabolism used by microbiota to survive and  
 785 divide in the gut may generate compounds, such as metabolic waste products, that affect host  
 786 behaviour as a side effect. In this case, the compounds are not adapted to influence the host, and  
 787 host effects are a by-product. Effects driven by natural selection on the host are shown in purple.  
 788 The host may evolve to depend on the microbiota for particular functions, including nutrient  
 789 provision or immune system maturation, such that a missing microbial species leads to strong  
 790 physiological effects and, potentially, behavioural effects. In addition, natural selection is  
 791 expected to favour hosts that use the microbiota to provide information on nutrition and health  
 792 in a manner that influences feeding, foraging and sickness behaviour. In all cases, the effects of  
 793 the microbiota may be due to multiple mechanisms, including the production of neuroactive  
 794 chemicals that then trigger the vagus nerve or travel to the brain through the blood or lymphatic  
 795 system, or through effects on the immune system.



814 **Figure 2. How neurotransmitters in the gut lumen might influence the central nervous system.**

815 Several neurotransmitters have been isolated from microbial species known to occur in the  
 816 human gut (examples in grey box)<sup>88</sup>. The microbial production of neurotransmitters represents a  
 817 potential mechanism to directly influence our brain and behaviour. In reality, this route is limited  
 818 because most neurotransmitters including serotonin, dopamine and GABA cannot typically  
 819 breach the protective blood-brain barrier<sup>81,130</sup> (1). Alternative modes of action include the  
 820 possibility that microorganism-derived neurotransmitters affect the brain through the vagus  
 821 nerve and its afferent neurones<sup>131</sup> (2). Another option is that precursors of neurotransmitters  
 822 cross the blood-brain barrier<sup>132,133</sup> and are then converted into active neurotransmitters (3). For  
 823 example, gut bacteria can influence the metabolism and availability of the serotonin precursor  
 824 tryptophan<sup>134</sup>. This may affect serotonergic signalling in the central nervous system as  
 825 tryptophan concentration in the blood plasma has been shown to correlate with brain serotonin  
 826 levels<sup>135</sup>.

827 **Table 1. Conditions favouring symbiont manipulation of a host.**

Evolutionary parameter <sup>a</sup>	Prediction	Parasite or pathogen example <sup>b</sup>	Hypothetical microbiota example	Likelihood for mammalian gut symbionts
<b>High benefit</b>	Host behaviour affects symbiont abundance within the host, and/or transmission.	<i>Ophiocordyceps unilateralis</i> fungus needs ant to move to specific elevation to develop <sup>33</sup> (Box 1).	Changes in host social interactions promote microbial transmission.	High
<b>Low cost</b>	Manipulation has limited negative effect on symbiont growth rate and survival, or manipulation is transient.	Nematomorph hairworms disperse by inducing their locust or grasshopper host to jump into water; involves only transient manipulation <sup>136</sup> .	Microbial waste product or signalling molecule happens to strongly affect host neurophysiology. Microorganism evolves manipulation by upregulating this pathway under specific conditions.	High
<b>High within-host abundance</b>	Abundant symbionts may benefit most if they can generate large amounts of manipulating compounds.	Many manipulative parasites reach high biomass within the host, for example <i>Ophiocordyceps unilateralis</i> (Box 1).	Highly abundant strain influences host behaviour. Bacteroidales strains reach high frequencies in the gut, although each is typically only a few percent of total microbial cells <sup>50</sup> .	Low
<b>Limited within-host evolution</b>	Symbiont undergoes few cell divisions within the host, either due to transient colonisation or occupying a slow-growing ecological niche.	<i>Salmonella enterica</i> serovar Typhimurium, which promotes host appetite (Box 1), only transiently infects the host <sup>52</sup> .	Microorganism is specialist on a low abundance nutrient in the gut.	Low
<b>Low genetic diversity</b>	Few other genotypes – mutants, strains or species – within the niche of the symbiont, which prevents a slow-growing manipulating strain being outcompeted.	<i>Wolbachia</i> strains have a diverse range of manipulative effects on insects and are intracellular, so little competition from other genotypes <sup>95</sup> .	Microorganism is in discrete compartment within host, limiting competition.	Low

828

829 a) Evolutionary theory predicts specific conditions that favour the persistence of a manipulating symbiont<sup>39–41</sup>. Not  
 830 all conditions are necessary for manipulation to evolve, for example, a symbiont that experiences little competition  
 831 (low genetic diversity) might evolve an energetically costly manipulation trait. Critically, however, theory predicts  
 832 that either limited within-host evolution or low genetic diversity is necessary for the evolution of manipulation (as  
 833 they prevent a non-manipulating strain from outcompeting a slower-growing manipulating strain). b) The best  
 834 candidate examples of host manipulation come from a few types of parasite or pathogen and we use these as  
 835 illustration. However, many parasites and pathogens do not appear to manipulate host behaviour. Indeed, some are  
 836 members of the microbiota with very similar ecologies to commensal and mutualistic microorganisms, making them  
 837 subject to the same constraints on the evolution of manipulation.

838

839

840

841 **Subject terms**

842 Biological sciences / Microbiology / Microbial communities / Microbiome

843 [URI /631/326/2565/2134]

844 Biological sciences / Microbiology / Microbial communities / Symbiosis

845 [URI /631/326/2565/547]

846 Biological sciences / Evolution / Coevolution

847 [URI /631/181/2481]

848 Biological sciences / Physiology / Neurophysiology

849 [URI /631/443/376]

850

851 **ToC blurb**

852 The microbiota can influence host behaviour through the gut-brain axis. In this Opinion, Johnson  
853 and Foster explore the evolution of this relationship and propose that adaptations of competing  
854 gut microorganisms may affect behaviour as a by-product, leading to host dependence.